Chiral Recognition of 1-(4-Quinolyl)ethanol by Permethylated α -Cyclodextrin

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¹H NMR studies on enantioselective complexation of 1-(4-quinolyl)ethanol (4QE) with hexakis(tri-O-methyl)- α cyclodextrin reveal the formation of the equatorial and axial inclusion complexes of (R) - and (S) -4QEs, respectively.

Cyclodextrins (CDs) have extensively been used as chiral selectors which can separate guest enantiomers in HPLC, GLC and capillary zone electrophoresis $(CZE)^1$ In spite of numerous examples of enantiomer separation by these analytical methods, CDs have generally been known as poor hosts to discriminate between optical isomers of guests in aqueous solutions.² Indeed, several studies show small differences in free energy changes $(\Delta \Delta G)$ for complexation between the enantiomers of guests having central chirality.⁸ Most of these studies, however, lack in clarification of the mechanisms for chiral recognition. In this work, we chose the 1-arylethanols, especially 4QE (see Fig. 1), as the typical guests with central chirality and studied the mechanism for

Fig. 1 Chiral 1-arylethanols used in the present study

Table 3 Binding constants (K) and enantioselectivities $(\Delta \Delta G)$ for complexation of (*R*)- and (*S*)-4QEs with various CDs in D₂O at pD 9.0 and 25 °C^a

^aThe K values were determined from ¹H NMR titrations of the enantiomers of 4QE (2 × 10⁻³ mol dm⁻³) in D₂O.
 ${}^b\Delta\Delta G = \Delta G_S - \Delta G_R$. ^cThe K values for the α -CD complexes could not be determined because of very small CIS. ^{*d*The $\Delta\Delta G$} values for these systems were not calculated because the differences in the K values between the enantiomers were in the range of the standard deviations.

their enantioselective complexation with CDs by means of ¹H NMR spectroscopy.

The binding constants (K) determined from the NMR titrations for complexation with hexakis(tri-O-methyl)-acyclodextrin (TMe- α -CD) are 51 \pm 3 and 41 \pm 1 dm³ mol⁻¹ for (R) - and (S) -40Es, respectively (Table 3). Similar enantioselective complexation was observed for 1NE and 1PyE whose molecular sizes are too large to be incorporated wholly into the TMe- α -CD cavity (Table 4). In contrast, no chiral discrimination between the enantiomers occurs with PhE, 3PE and 4PE whose molecular sizes are significantly smaller than those of 4QE, 1NE and 1PyE. The ROESY spectra as well as the molecular mechanics-molecular dynamics calculations suggest the formation of the equatorial and axial inclusion complexes of (R) - and (S) -4QEs, respectively (Fig. 7). The chiral discrimination might occur at the secondary OCH₃ group side of TMe- α -CD where the guest molecule is bound shallowly to the asymmetrically twisted cavity of the host.

Table 4 Binding constants (K) and enantioselectivities $(\Delta \Delta G)$ for complexation of (R) - and (S) -1-arylethanols with TMe- α -CD in D₂O at pD 9.0 and 25 °C

Guest	$K_1/$ dm^3 mol ⁻¹	$\frac{K_2}{\text{dm}^3}$ mol ⁻¹	$\Delta\Delta G/$ kJ mol ^{-1b}
(R) - and (S) -PhE (R) - and (S) -4PE	$128 + 22$ $34 + 2$		0 0
$(R) - 1NE$ $(S) - 1NE$	$277 + 14$ $239 + 15$	$41 + 3$ $28 + 5$	$0.37 + 0.28$
$(R) - 4QE$ $(S) - 4QE$ $(R) - 1$ PyE	$51 + 3$ $41 + 1$ 565 ± 57		$0.54 + 0.20$
$(S) - 1$ PyE	$421 + 43$		$0.73 + 0.51$

^aThe K values were determined from ¹H NMR titrations of 1-arylethanols $(2 \times 10^{-3} \text{ mol dm}^{-3})$ in D₂O except for 1PyE. In the case of 1PyE, the K values were determined by means of
UV–VIS absorption spectroscopy in 0.033 mol dm⁻³ phosphate buffer at pH 9.0. ${}^b\Delta\Delta G = \Delta G_S - \Delta G_R$. The $\Delta\Delta G$ for K_1 is *To receive any correspondence. exhibited for the 1NE complexes.

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Fig. 7 Structures of the (R) - and (S) -40E complexes of TMe- α -CD deduced from the ROESY spectral measurements

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Techniques used: ¹H NMR, molecular dynamics-molecular mechanics calculations

Appendix: NMR titration curves for determining the K values

Table 1: Results on ¹H NMR measurements of 1-arylethanols $(2 \times 10^{-3} \text{ mol dm}^{-3})$ in D₂O at pD 9.0 in the presence of various CDs at 25° C

Table 2: Results on ¹H NMR measurements of 1-arylethanols $(2 \times 10^{-3} \text{ mol dm}^{-3})$ in D₂O at pD 9.0 in the presence of TMe- α -CD $(1 \times 10^{-2} \text{ mol dm}^{-3})$ at 25 °C

Fig. 2: ¹H NMR spectra of (\pm)-4QE (2×10^{-3} mol dm⁻³) in D₂O at pD 9.0 in the absence and the presence of various CDs at 25 °C

Fig. 3: ¹H NMR spectra of (\pm) -9AnE $(2 \times 10^{-3} \text{ mol dm}^{-3})$ in CDCl₃ and D₂O at pD 9.0 in the presence of TMe- α -CD $(1 \times 10^{-2} \text{ mol dm}^{-3})$ at 25° C

Fig. 4: Changes in the chemical shifts of TMe- α -CD $(2 \times 10^{-3}$ mol dm⁻³) in D₂O at pD 9.0 and 25 °C upon addition of (R) - and (S)-4QEs and 1NEs $(2 \times 10^{-3} \text{ mol dm}^{-3})$

Fig. 5: ROESY spectrum of the (R) -40E-TMe- α -CD system at 25 °C. The sample was the mixture of (R) -4QE $(4 \times 10^{-3} \text{ mol dm}^{-3})$ and TMe- α -CD $(1 \times 10^{-2} \text{ mol dm}^{-3})$ in D₂O at pD 9.0 and 25 °C

Fig. 6: ROESY spectrum of the (S) -4QE-TMe- α -CD system at 25 °C. The sample was the mixture of (S) -4QE $(4 \times 10^{-3} \text{ mol dm}^{-3})$ and TMe- α -CD (1 × 10⁻² mol dm⁻³) in D₂O at pD 9.0 and 25 °C

Fig. 8: Structures of the (R) -40E $-$ and (S) -40E $-TMe-_{\alpha}-CD$ complexes in water calculated from the MM-MD calculations

References: 20

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References cited in this synopsis

- 1 (a) V. Schurig and H.-P. Nowotny, Angew. Chem., Int. Ed. Engl., 1990, 29, 939; (b) S. Li and W. C. Purdy, Chem. Rev., 1992, 92, 1457; (c) S. F. Y. Li, J. Chromatogr. Library, 1992, 52, 201; (d) R. Kuhn and S. Hoffstetter-Kuhn, Chromatographia, 1992, 34, 505; (e) S. Fanali, J. Chromatogr. A, 1996, 735, 77.
- 2 For reviews, see (a) K. Kano, in Bioorganic Chemistry Frontiers, ed. H. Dugas and F. P. Schmidtchen, Springer-Verlag, Berlin, Heidelberg, 1993, vol. 3, ch. 1; (b) K. Kano, J. Phys. Org. Chem., 1997, 10, 286.
- (a) F. Cramer and W. Dietsche, Chem. Ber., 1959, 92, 378; (b) A. Cooper and D. D. MacNicol, J. Chem. Soc., Perkin Trans. 2, 1978, 760; (c) Y. Ihara, E. Nakanishi, M. Nango and J. Koga, Bull. Chem. Soc. Jpn., 1986, 59, 1901; (d) Y. Inoue, F.-H. Kuan and R. Chujo, Bull. Chem. Soc. Jpn., 1987, 60, 2539; (e) M. Barra and R. H. de Rossi, J. Org. Chem., 1989, 54 , 5020 ; (f) S. E. Brown, J. H. Coates, S. F. Lincoln, D. R. Coghlan and C. J. Easton, J. Chem. Soc., Faraday Trans., 1991, 87, 2699; (g) S. Li and W. C. Purdy, Anal. Chem., 1992, 64, 1405; (h) Y. Yamashoji, T. Ariga, S. Asano and M. Tanaka, Anal. Chim. Acta, 1992, 268, 39; (i) J. B. Cunniff and P. Vouros, J. Am. Mass Spectrom., 1995, 6, 437.